

Sex Differences in the Neuroadaptations of Reward-related Circuits in Response to Subchronic Variable Stress

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Abstract—Women are twice as likely to be diagnosed with major depressive disorder. However, fewer studies in rodent models of depression have used female animals, leading to a relative lack of understanding of the female brain's response to stress, especially at a neural circuit level. In this study, we utilized a 6-day subchronic variable stress (SCVS) mouse model and measured novelty suppressed feeding as behavioral criteria to evaluate susceptibility to SCVS in male and female mice. First, we showed that SCVS induced a decrease in latency to eat (susceptible phenotype) in female mice, but not in males (resilient phenotype). After determining behavioral phenotypes, we investigated the firing activities of dopamine (DA) neurons in the ventral tegmental area (VTA), as well as the neurons that project from lateral habenula (LHb) to the VTA and from locus coeruleus (LC) to the VTA. Utilizing retrograding lumafuor fluorescent tracers and electrophysiology techniques, we performed cell type- and circuit-specific measures of neuronal firing rates. Our data show that SCVS significantly increased the firing rate of LHb-VTA circuit neurons in female mice when compared to that of their female controls, an effect that was absent in SCVS-exposed males. Interestingly, SCVS did not induce significant firing alterations in VTA DA neurons and LC-VTA circuit neurons in either female mice or male mice when compared to their stress-naïve controls. Overall, our data show sex differences in the LHb-VTA circuit responses to SCVS, and implicates a potential role of this projection in mediating vulnerability of female mice to stress-induced depression. © 2018 IBRO. Published by Elsevier Ltd. All rights reserved.

Key words: sex difference, major depression, neuronal activity, ventral tegmental area, lateral habenula, locus coeruleus.

INTRODUCTION

According to a WHO report (Piccinelli and Gomez Homen, 1997), women are twice as likely as men to be diagnosed with stress-related psychiatric disorders, including major depression and anxiety disorder (Kessler et al., 1994; Kendler et al., 1995; Kessler,

2003). It has been suggested that females may respond differently to stress and use distinct stress-coping strategies as compared to males (Kendler et al., 2001; Maciejewski et al., 2001; Klein and Corwin, 2002; Nemeroff et al., 2006). While widely used rodent models of depression have contributed enormously to unravel the neural mechanisms that underlie behavioral responses to stress, these animal models have largely used males only (Solomon et al., 2007; Dalla et al., 2008; Trainor et al., 2011; Ver Hoeve et al., 2013). Due to these limitations, the mechanisms underlying sex differences in stress vulnerability remain largely unknown.

The subchronic variable stress (SCVS) model is used to cause stress-induced depression by exposing mice to three alternating stressors across 6 days (LaPlant et al., 2009; Hodes et al., 2015). Historically, this paradigm induces a depressive phenotype in females, but not in

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Abbreviations: aCSF, artificial cerebrospinal fluid; CMS, chronic mild stress; CRF, corticotropin-releasing factor; DA, dopamine; DBS, deep-brain stimulation; LC, locus coeruleus; LH, learned helplessness; LHb, lateral habenula; MHb, medial habenula; NAc, nucleus accumbens; NE, norepinephrine; NSF, novelty suppressed feeding; RSDS, repeated social defeat stress; SCVS, subchronic variable stress; VTA, ventral tegmental area.

39 males. Thus, the SCVS model provides an ideal model for
 40 exploring sex differences of vulnerability to stress. Based
 41 on this model, it has also been demonstrated that male
 42 and female mice show differential patterns of gene
 43 expression in the nucleus accumbens (NAc) (Hodes
 44 et al., 2015). Furthermore, Dnmt3a and NF- κ B have been
 45 identified as important mediators that contribute to sex dif-
 46 ferences between male and female mice in stress sus-
 47 ceptibility (LaPlant et al., 2009; Hodes et al., 2015). In
 48 contrast, less is known about the neurophysiological
 49 mechanisms that underlie sex differences in stress
 50 vulnerability.

51 The ventral tegmental area (VTA) dopamine (DA)
 52 system is a key part of the brain's reward circuitry and
 53 plays an important role in mediating stress response
 54 (Chaudhury et al., 2013; Friedman et al., 2014). The lat-
 55 eral habenula (LHb) and the locus coeruleus (LC) both
 56 send substantial efferents to innervate VTA DA neurons
 57 (Omelchenko et al., 2009; Chandler et al., 2013; Juarez
 58 and Han, 2016). Accordingly, multiple lines of evidence
 59 have demonstrated that the VTA, LHb and LC brain areas
 60 all play crucial roles in mediating stress responses. The
 61 firing activities of neurons in these three brain regions
 62 are altered when exposed to various stressors (Ungless
 63 et al., 2004; Krishnan et al., 2007; Sartorius and Henn,
 64 2007; Cao et al., 2010; Li et al., 2011, 2013; Valenti
 65 et al., 2012; Isingrini et al., 2016). Furthermore, recent
 66 studies show that the firing activities of VTA DA neurons,
 67 LHb-VTA circuit neurons and LC-VTA circuit neurons
 68 determine susceptible versus resilient behaviors seen in
 69 male animals in a variety of stress-induced psychiatric
 70 disorders (Li et al., 2011; Chaudhury et al., 2013; Tye
 71 et al., 2013; Isingrini et al., 2016). However, how the activ-
 72 ity of these neurons is affected following the SCVS para-
 73 digm remains unknown in both female and male mice.

74 Here, utilizing the SCVS model and cell type- and
 75 projection-specific *in vitro* electrophysiological recording
 76 techniques, we investigated the firing activity alterations
 77 of VTA DA neurons, LHb-VTA projecting neurons and
 78 LC-VTA projecting neurons in both male and female
 79 mice following SCVS. Our findings provide useful
 80 evidence that the hyper-activation of the LHb-VTA
 81 circuit may play an important role in mediating the
 82 vulnerability of female mice to stress-related disorders,
 83 as compared to males.

84 EXPERIMENTAL PROCEDURES

85 Animals

86 Seven-week-old C57BL/6J female and male mice (The
 87 Jackson Laboratory) were used to set up the SCVS
 88 paradigm. All mice were group-housed on a 12-h
 89 light/dark cycle with food and water available *ad libitum*.
 90 Following the last day of SCVS, all mice were singly
 91 housed for behavioral testing and *in vitro* recording. All
 92 procedures were approved by the Institutional Animal
 93 Care and Use Committee of the Icahn School of
 94 Medicine at Mount Sinai and in accordance with the
 95 National Institutes of Health guidelines. Twenty-eight
 96 female mice and twenty-five male mice were used in
 97 this study. Electrophysiological recordings were

obtained from the same cohort of mice after behavioral 98
 tests (see below). 99

100 Subchronic variable stress

SCVS was performed as described previously (Hodes 101
 et al., 2015). Female and male mice were put through 102
 three unpredictable stressors over 6 days (Fig. 1A). To 103
 prevent habituation, mice were subjected to stress in 104
 the following order: 100 random foot shocks at 0.45 mA 105
 for 1 h (6–8 mice/chamber) on day 1 and day 4; tail sus- 106
 pension stress in which all mice were fixed to hang in 107
 an inverted position for 1 h on day 2 and day 5; and 108
 restraint stress, in which mice were placed inside a 50- 109
 ml falcon tube for 1 h within the home cage on day 3 110
 and day 6. After each stressor, mice were returned to their 111
 home cage except on the last day of SCVS when they 112
 were then singly housed. 113

114 Novelty suppressed feeding (NSF)

115 It has been shown that the SCVS paradigm induces 115
 several consistent behavioral deficits in female mice, but 116
 not in males, specifically depression-associated 117
 behaviors (Hodes et al., 2015). To minimize the effect of 118
 behavioral test-related stress on our electrophysiological 119
 recordings, we chose less stressful NSF as a test to con- 120
 firm the behavioral phenotypes before carrying out the 121
 electrophysiological experiments. The NSF test was car- 122
 ried out as previously described (Santarelli et al., 2003; 123
 Hodes et al., 2015). Briefly, mice were food deprived 24 124
 h before testing. Water was offered *ad libitum*. On the 125
 day of testing, mice were transferred to the testing room 126
 1 h prior to start of the experiment. Under red light condi- 127
 tions, corncob bedding was lightly distributed on the floor 128
 of a plastic box of 50 × 50 × 20 cm. A single food pellet 129
 was placed on a platform where a petri dish was covered 130
 with a white circle cut out from Whatman paper and the 131
 platform was positioned in the center of the box. Mice 132
 were then placed in the corner of the box and a timer 133
 was started. The latency for mice grasping the food pellet 134
 with their forepaws and biting was recorded with a limit up 135
 to 10 min during testing. As soon as the mice began to 136
 eat, or the 10-min time limit was reached, they were 137
 immediately transferred back to their home cage. 138

139 *In vitro* slice electrophysiology

140 The electrophysiological recording procedures were 140
 followed as previously described (Chaudhury et al., 141
 2013; Friedman et al., 2014). Under blinded conditions, 142
 mice were anesthetized with isoflurane and perfused 143
 immediately for 40–60 s with ice-cold artificial cere- 144
 brospinal fluid (aCSF) containing (in mM): 128 NaCl, 3 145
 KCl, 1.25 NaH₂PO₄, 10 D-glucose, 24 NaHCO₃, 2 CaCl₂ 146
 and 2 MgCl₂ (oxygenated with 95% O₂ and 5% CO₂, 147
 pH 7.4, 295–305 mOsm). Acute brain slices (250 μ m) 148
 containing VTA, LHb or LC were cut using a vibratome 149
 microslicer (DTK-1000, Ted Pella) in ice-cold sucrose 150
 aCSF, which was derived by fully replacing NaCl with 151
 254 mM sucrose and saturated by 95% O₂ and 5% 152
 CO₂. Slices were maintained in holding chambers with 153

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