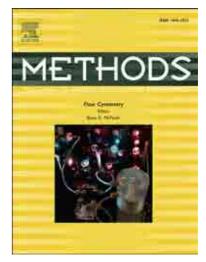
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Dissecting single–cell molecular spatiotemporal mobility and clustering at Focal Adhesions in polarised cells by fluorescence fluctuation spectroscopy methods

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- 1 Dissecting single-cell molecular spatiotemporal mobility and clustering at Focal Adhesions in
- 2 polarised cells by fluorescence fluctuation spectroscopy methods
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- 8

9 Abstract

Quantitative fluorescence fluctuation spectroscopy from optical microscopy datasets is a very 10 powerful tool to resolve multiple spatiotemporal cellular and subcellular processes at the molecular 11 12 level. In particular, raster image correlation spectroscopy (RICS) and number and brightness analyses (N&B) yield molecular mobility and clustering dynamic information extracted from real-time cellular 13 14 processes. This quantitative information can be inferred in a highly flexible and detailed manner, i.e. 15 1) at the localisation level: from full-frame datasets and multiple regions of interest within; and 2) at the temporal level: not only from full-frame and multiple regions, but also intermediate temporal 16 17 events. Here we build on previous research in deciphering the molecular dynamics of paxillin, a main 18 component of focal adhesions. Cells use focal adhesions to attach to the extracellular matrix and 19 interact with their local environment. Through focal adhesions and other adhesion structures, cells 20 sense their local environment and respond accordingly; due to this continuous communication, these structures can be highly dynamic depending on the extracellular characteristics. By using a 21 22 previously well-characterised model like paxillin, we examine the powerful sensitivity and some 23 limitations of RICS and N&B analyses. We show that cells upon contact to different surfaces show 24 differential self-assembly dynamics in terms of molecular diffusion and oligomerisation. In addition, 25 single-cell studies show that these dynamics change gradually following an antero-posterior 26 gradient.

27

28 Keywords: Fluorescent Fluctuation Spectroscopy; Molecular Brightness; Protein dynamics; Protein

29 clustering; Focal adhesions; Cell Biophysics.

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